

# **INTERPRETATION AND USE OF SILAGE FERMENTATION ANALYSIS REPORTS**

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## **INTRODUCTION**

Fermentation analyses have long been used in university and industry research trials to assess silage quality. These analyses are now available for evaluating silage quality on farms through commercial forage testing laboratories. Analyses commonly included in silage fermentation reports are pH, lactic, acetic, propionic and butyric acids, ammonia, and ethanol. The purpose of this paper will be to discuss the usefulness of these reports.

## **SILAGE FERMENTATION ANALYSES**

Collectively, data from a fermentation analyses can tell us whether an excellent, average, or poor fermentation has occurred. Based on these analyses, we can usually make some educated assumptions about the kinds of microorganisms that controlled the ensiling process. In many, but not all cases, the fermentation that a crop undergoes can be explained by various crop factors such as moisture content, buffering capacity, and sugar content. However, management factors such as silo packing speed, silage pack density, type of additive used, chop length, silo management during storage, and silo management during feed-out can affect fermentation analyses. In some cases fermentation analyses can qualitatively explain poor silage nutritive value or low intakes, but they cannot be used to balance diets for cattle. Thus, they should always be used in conjunction with other standard chemical analyses (i.e. ADF, NDF, CP, RDP/RUP, NE<sub>L</sub>, NDF digestion, etc.).

## RANGES FOR SILAGE FERMENTATION ANALYSES

Table 1. Concentrations of common fermentation end products in various silages.

Item	Legume Silage, 30 - 40% DM <sup>1</sup>	Legume Silage, 45 - 55% DM	Grass Silage, 30-35% DM	Corn Silage, 30-40% DM	HM Corn, 70-75% DM
pH	4.3 - 4.7	4.7 - 5.0	4.3 - 4.7	3.7 - 4.2	4.0 - 4.5
Lactic acid, %	7 - 8	2 - 4	6 - 10	4 - 7	0.5 - 2.0
Acetic acid, %	2 - 3	0.5 - 2.0	1 - 3	1 - 3	< 0.5
Propionic acid, %	< 0.5	< 0.1	< 0.1	< 0.1	< 0.1
Butyric acid, %	< 0.5	0	0.5 - 1.0	0	0
Ethanol, %	0.2 - 1.0	0.5	0.5 - 1.0	1 - 3	0.2 - 2.0
Ammonia- N, % of CP	10 - 15	< 12	8 - 12	5 - 7	< 10

<sup>1</sup>DM = Dry matter.

### SILAGE pH

The pH of an ensiled sample is a measure of its acidity, but is also affected by the buffering capacity of the crop. Two samples may have the same pH, but different concentrations of acids. In general, legume silages have a higher pH than corn or other grass silages and take longer to ensile because of their higher buffering capacity. Seldom do corn silages have a pH higher than 4.2. Such cases may be associated with extremely dry (>42% dry matter) silages that are overly mature or drought stricken. Because of its normally low pH (3.8), corn silage intake usually benefits from the addition of sodium bicarbonate prior to feeding to neutralize its acidity.

Common reasons for legume silages having a pH higher than 4.6 to 4.8 include: ensiling at <30% dry matter (DM) which causes a clostridial fermentation, and ensiling at > 45-50% DM, which restricts fermentation. In the first example, a high pH due to clostridia is a definite indicator of an undesirable fermentation that has led to poor quality silage. However, in the second example, a high pH due to restricted fermentation is not always indicative of a poor fermentation or poor silage. But, silage from a restricted fermentation usually is unstable when exposed to air because insufficient amounts of acid were produced to inhibit secondary microbial growth.

Some common reasons for a high silage pH are as follows:

- dry silage (> 50% DM)
- silage not fully fermented due to early sampling time relative to harvest, cold weather during harvest, and slow or poor packing
- legume silages with extremely high ash contents (> 15% of DM) and (or) high protein content (> 23-24% CP)
- silage with excess ammonia or urea
- clostridial silages
- spoiled or moldy silages
- silages containing manure.

### **BUFFERING CAPACITY**

Buffering capacity measures to what degree a forage sample will resist a change in pH. All forages have different buffering capacities. A fresh forage with a high buffering capacity will require more acid to reduce its pH than a forage with a low buffering capacity. In general, fresh legumes have a higher buffering capacity than do fresh grasses or corn.

### **LACTIC ACID**

Lactic acid should be the primary acid in good silages. This acid is stronger than other acids in silage (acetic, propionic and butyric) and thus usually responsible for most of the drop in silage pH. Fermentations that produce lactic acid result in the fewest losses of dry matter and energy from the crop during storage.

Some common reasons for low lactic acid content are as follows:

- restricted fermentation due to high DM content (especially legumes and grasses with > 50% DM).
- restricted fermentation due to cold weather.
- sample taken after considerable aerobic exposure that has degraded lactic acid.
- silages high in butyric acid (Clostridial silages) are usually low in lactic acid.

Lactic acid should be at least 65 to 70% of the total silage acids in good silage.

### **ACETIC ACID**

Extremely wet silages (< 25% DM), prolonged fermentations (due to high buffering capacity), loose packing, or slow silo filling can result in silages with high concentrations of acetic acid (>3 to 4% of DM). In such silages, energy and DM recovery are probably less than ideal. Silages treated with ammonia also tend to have higher concentrations of acetic acid than untreated silage, because the fermentation is prolonged by the addition of the ammonia that raises pH.

The effect of high concentrations of acetic acid (> 4 to 6% of DM) in silages fed to animals is unclear at this time. In the past, some studies can be found where DM intake was depressed when silage high in acetic acid concentration was fed to ruminants. However, the depression in intake to high acetic acid in the diet has not been consistent. There has been speculation that decreased intake may be actually due to unidentified negative factors associated with a poor fermentation and not to acetic acid itself. A new microbial inoculant (*Lactobacillus buchneri*) designed for improving the aerobic stability of silages causes higher than normal concentrations of acetic acid in silages. However, production of acetic acid from this organism should not be mistaken for a poor fermentation and feeding treated silages with a high concentration of acetic acid does not appear to cause negative effects on animal intake.

If a producer has intake problems due to silages with excessively high acetic acid (> 5 to 6% of DM), the amount of that silage should be reduced in the TMR. Other alternatives for managing these silages include: aerating the silage for a day to volatilize the acetic acid, removing the silage and then gradually reincorporating it back into the diet over a 2–3 week period, and partially neutralizing the silage with sodium bicarbonate prior to feeding (about 0.5 to 1% addition on DM basis).

### **PROPIONIC ACID**

Most silages contain very low concentrations of propionic acid (< 0.2 to 0.3%) unless the silage is very wet (< 25% DM). In silages with more typical concentrations of DM (35 to 45% DM), concentrations of propionic acid may be undetectable.

Biological additives that theoretically increase the propionic acid concentration of silage usually contain bacteria from the Propionibacteria family. However, research suggests that these organisms are usually unable to compete in normal silage environments and are thus, usually ineffective.

Chemical additives containing propionic acid are more effective for increasing the concentration of this acid in silages. These additives can range markedly in their percentage of active ingredients but most mainstream products will increase the concentration of propionic acid at ensiling from 0 to about 0.15 to 0.30% (DM basis) if added at 2 to 4 lb per ton of wet (~35% DM) silage.

### **BUTYRIC ACID**

A high concentration of butyric acid (>0.5% of DM) indicates that the silage has undergone clostridial fermentation, which is one of the poorest fermentations. Silages high in butyric acid are usually low in nutritive value and have higher ADF and NDF levels because many of the soluble nutrients have been degraded. Such silages may also be high in concentrations of soluble proteins and may contain small protein compounds called amines that have sometimes shown to adversely affect animal performance.

High concentrations of butyric acid has sometimes induced ketosis in lactating cows and because the energy value of silage is low, intake and production can suffer. As with other poor quality silages, total removal or dilution of the poor silage is advised.

### **AMMONIA**

High concentrations of ammonia (>10 to 15% of CP) are a result of excessive protein breakdown in the silo caused by a slow drop in pH or clostridial action. In general, wetter silages have higher concentrations of ammonia. Extremely wet silages (< 30% DM) have even higher ammonia concentrations because of the potential for clostridial fermentation. Silages packed too loosely and filled too slowly also tend to have high ammonia concentrations.

Theoretically, high amounts of ammonia (by itself) in silage should not have negative effects on animal performance if the total dietary nitrogen fractions are in balance. However, if the high ammonia contributes to an excess of ruminally-degraded protein (RDP), this could have negative consequences on milk and reproductive performances. Blood or milk urea nitrogen can be used as an indicator of excess RDP. Often times, silage with high concentrations of ammonia coupled with butyric acid may also have significant concentrations of other undesirable end products, such as amines, that may reduce animal performance.

### **ETHANOL**

High concentrations of ethanol are usually an indicator of excessive metabolism by yeasts. Dry matter recovery is usually worse in silages with large numbers of yeasts. These silages are also usually very prone to spoilage when the silage is exposed to air. Usual amounts of ethanol in silages are low (< 1 to 2% of DM). We do not know the level at which ethanol becomes a problem in dairy cattle diets. Most ethanol that is consumed is probably converted to acetic acid in the rumen. Extremely high amounts of ethanol (> 3 to 4% of DM) in silages may cause off flavors in milk.

### **USING FERMENTATION ANALYSES**

Assuming that all nutrients are balanced properly in the diet, silage fermentation reports may be able to identify the reason for intake or performance problems. However, they cannot be used to balance rations.

It is extremely difficult to use a fermentation analysis to determine whether a silage additive was used unless done in a well controlled research trial. However, one might be able to tell if a propionic acid product were added to silage since most silages typically are low in this acid. If silage was treated with about 2 lb/ton of fresh forage using a buffered propionic acid product, one might be able to detect the increase in this acid. Most silages have < 0.15% propionic acid.

## **SAMPLING OF SILAGES**

If your goal is to evaluate what animals are being fed, then samples should be reflective of this by sampling at feeding. If your goal is to see what type of fermentation process the silage went through, then collect a sample that is as fresh as possible and that has not been exposed to air (typically at least 8 to 10 inches below or beyond the face).

For chemical fermentation analyses, samples should be frozen immediately after being taken, then shipped preferably on ice in a cooler by a next day delivery service. Ship at the beginning, not end of the week so samples will not be held up during the weekend.

## **COMMERCIAL LABS CONDUCTING SILAGE ANALYSES**

To the best of our knowledge, as of August 2001, the following commercial forage testing laboratories perform fermentation analyses:

- Dairy One  
730 Warren Road  
Ithaca, NY 14850  
Contact: Paul Sirois, 800 496 3344, [www.DairyOne.com](http://www.DairyOne.com)
- Cumberland Valley Analytical Services  
Maugansville, MD  
Contact: Ralph Ward, (301) 790-1980, [cvas@intepid.net](mailto:cvas@intepid.net)
- Near-infrared  
Rock River Laboratories  
Watertown, WI  
Contact: Don Meyer, (920) 261-0446, [rllab@execpc.com](mailto:rllab@execpc.com)
- High-performance liquid chromatography  
Dairyland Laboratories  
Arcadia, WI  
Contact: Dave Taysom, (608) 323-2123, [dtaysom@dairylandlabs.com](mailto:dtaysom@dairylandlabs.com)

Current information on analytical costs and sample turn-around time is best obtained directly from the individual labs.